

TEN TIPS

FOR WORKING IN YOUR NUAIRE BIOLOGICAL SAFETY CABINET

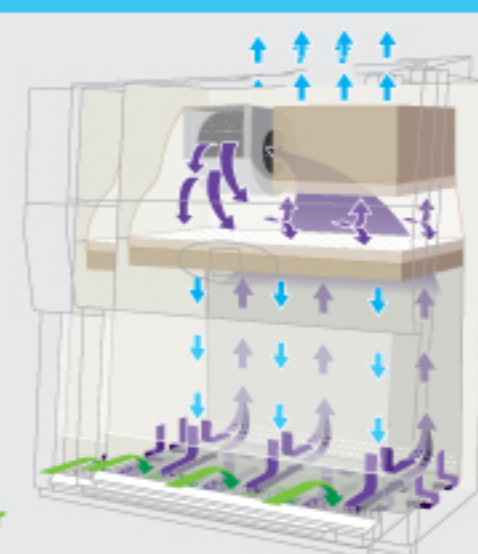


Good technique when working within a Class II Biological Safety Cabinet (BSC) will minimize air turbulence and prevent splatter or unwanted spread of aerosols. Here are some tips for good technique that will maximize potential protection of personnel, product and environment.

TIP 1: KNOW YOUR AIRFLOW

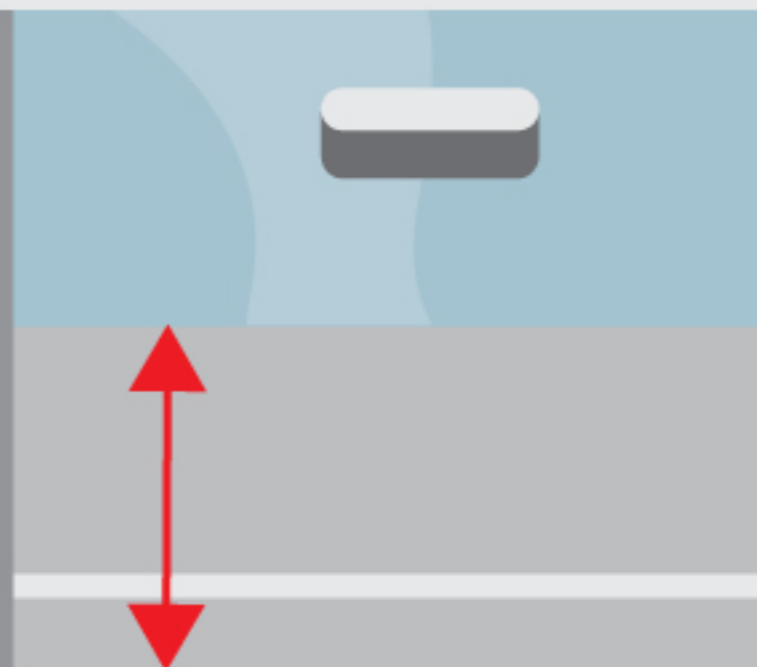
Biological Safety Cabinets provide personnel, product, and environmental protection through the use of HEPA filtered air. Knowing how the air is passed through the cabinet an essential piece of the puzzle to know

→ HEPA Filtered Air → Contaminated Worksurface Air → Contaminated Room Air



TIP 2: WORK AT THE PROPER SASH LEVEL

Proper airflows are reached by a certifier measuring air speed on an annual basis (or more) to put a stamp of approval that the cabinet is fully operation to provide personnel, product, and environmental protection



TIP 3: NEVER COVER THE AIR GRILL

Covering the air grill at the front of the cabinet compromises airflow integrity. Blocking the grill can cause laboratory air to enter the work zone or even work zone air to enter the laboratory environment.

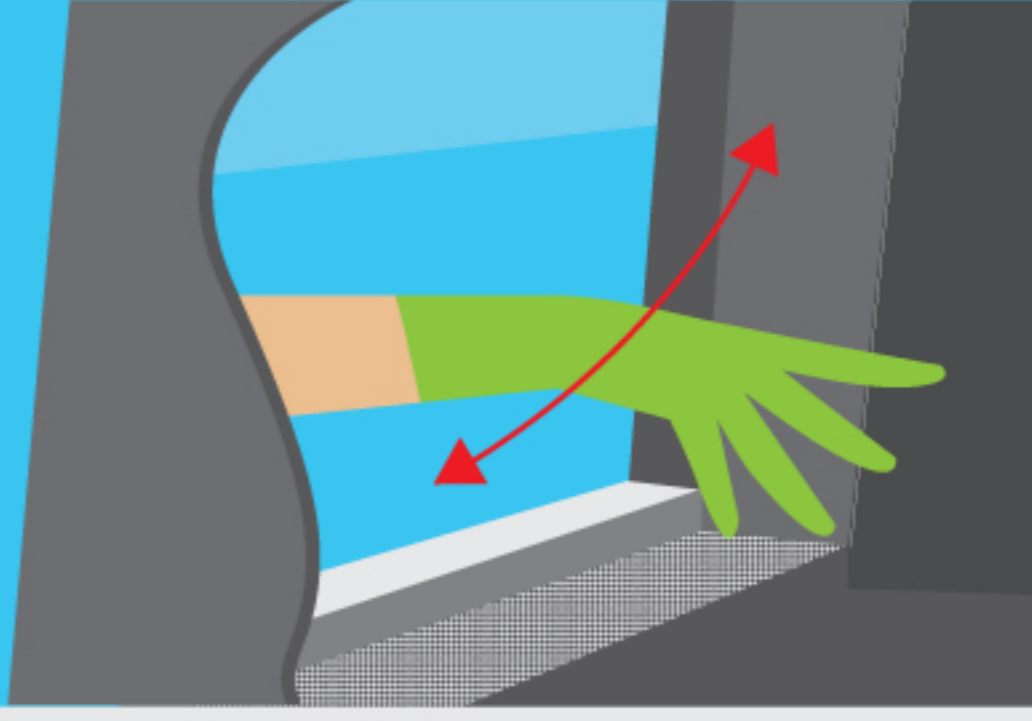
PS. This goes for your elbows and arms as well. NuAire offers elbow rest accessories to avoid this occurrence.



TIP 4: MINIMIZE MOVEMENT

The rapid movement of a worker's arms in a sweeping motion into and out of the cabinet will disrupt the air curtain and may compromise the partial barrier containment provided by the BSC.

Moving arms in and out slowly, perpendicular to the face opening of the cabinet, will reduce this risk. Other personnel activities in the room (e.g., rapid movement, open/closing room doors, etc.) may also disrupt the cabinet air barrier.

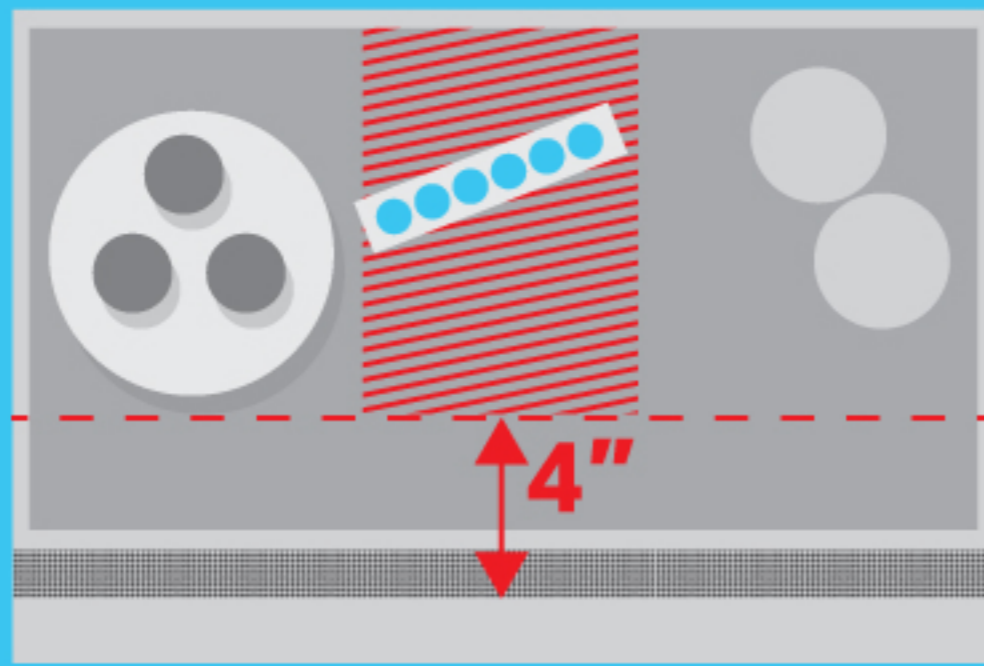


TIP 5: REDUCE SPLATTER

Many common procedures conducted in Biological Safety Cabinets may create splatter or aerosols. Good microbiological techniques should always be used when working in a Biological Safety Cabinet to minimize this potential.

For example, techniques to reduce splatter and aerosol generation will minimize the potential for personnel exposure to infectious materials manipulated within the cabinet.

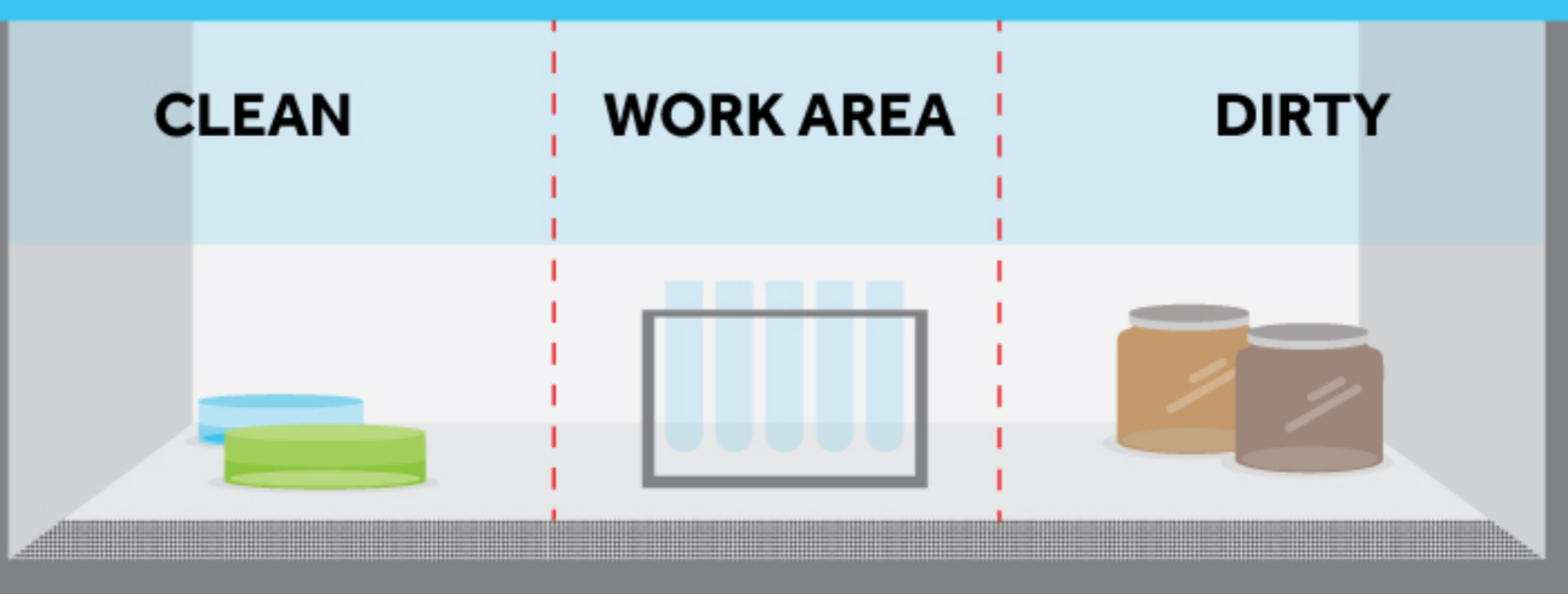
Class II cabinets are designed so that horizontally nebulized spores will be captured by the downward flowing cabinet air within fourteen inches (35cm) of travel. As a general rule of thumb, keeping clean materials at least twelve inches (31cm) away from aerosol-generating activities will minimize the potential for cross-contamination.



TIP 6: KNOW YOUR WORK AREA

The middle third of the work surface is the ideal area to be used. All operations should be performed at least 4 inches from the front grille on the work surface.

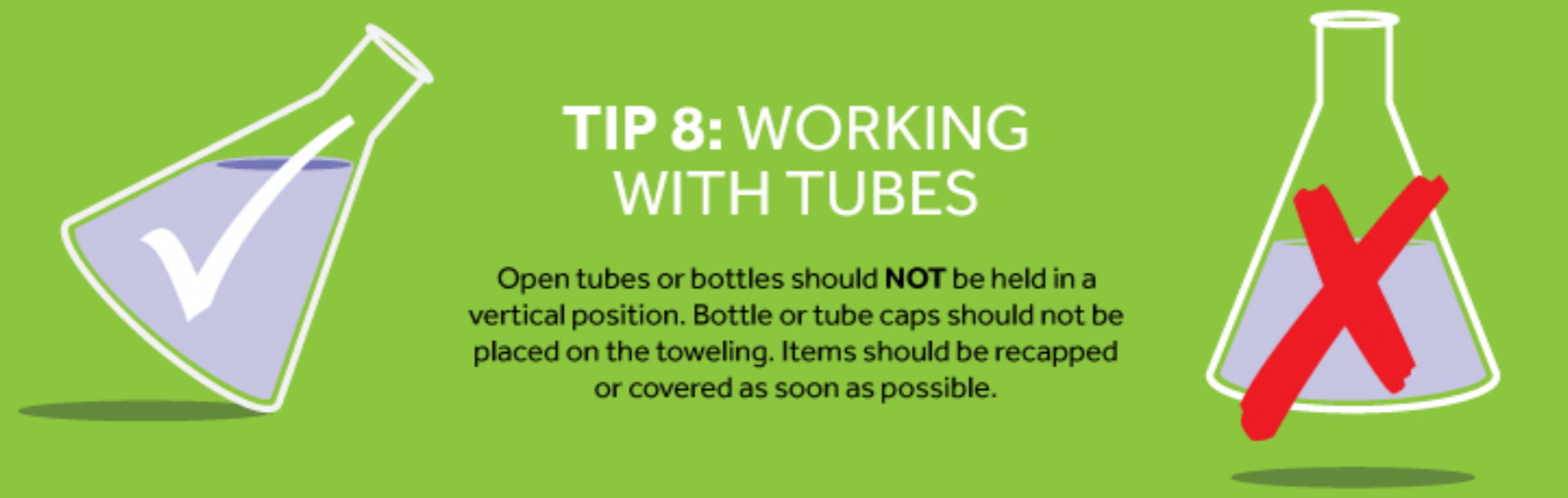
Materials or equipment placed inside the cabinet may cause disruption to the airflow, resulting in turbulence, possible cross-contamination, and/or breach of containment.



TIP 7: WORK FROM CLEAN TO CONTAMINATED

Active work should flow from the clean to contaminated area across the work surface. Materials and supplies should be placed in such a way as to limit the movement of "dirty" items over "clean" ones.

Maintain proper balance of materials from left to right in order to prevent an airflow imbalance within the work zone. Bulky items such as biohazard bags, discard pipette trays and suction collection flasks should be placed to one side of the interior of the cabinet. All materials should be placed as far back in the cabinet as practical, toward the rear edge of the work surface and away from the front grille of the cabinet. Similarly, aerosol-generating equipment (e.g., vortex mixers, tabletop centrifuges) should be placed toward the rear of the cabinet to take advantage of the air split.

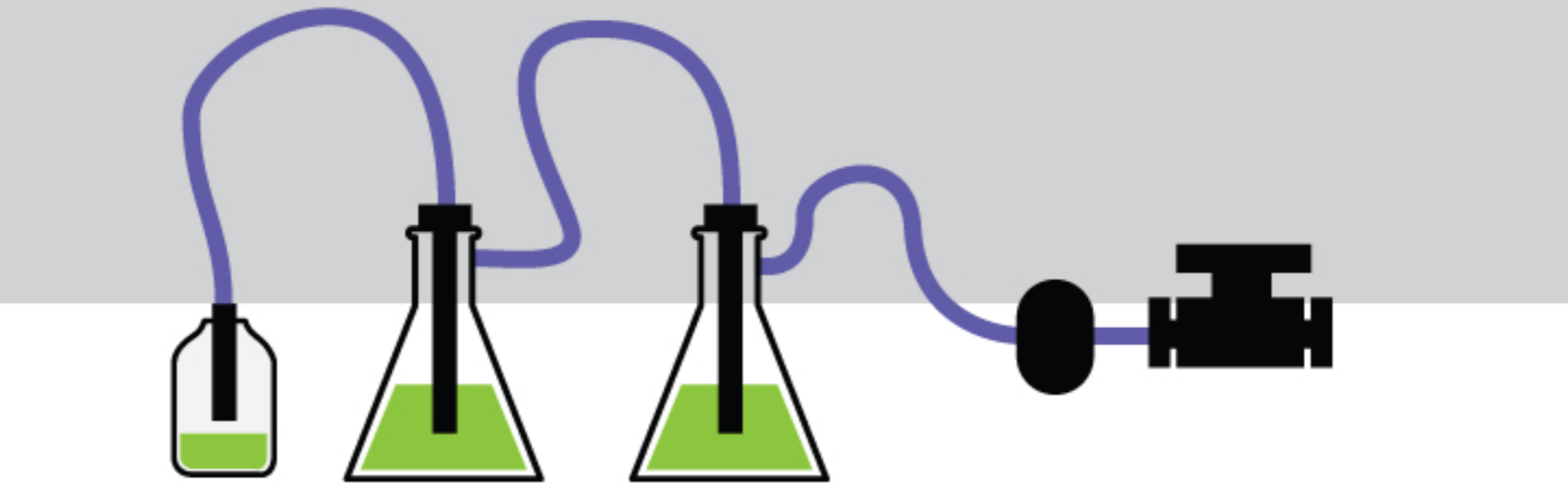


TIP 8: WORKING WITH TUBES

Open tubes or bottles should NOT be held in a vertical position. Bottle or tube caps should not be placed on the towel. Items should be recapped or covered as soon as possible.

TIP 9: WORKING WITH PETRI DISHES

Investigators working with Petri dishes and tissue culture plates should hold the lid above the open sterile surface to minimize direct impaction of downward air.



TIP 10: WORKING WITH ASPIRATOR BOTTLES OR SUCTION FLASKS

Aspirator bottles or suction flasks should be connected to an overflow collection flask containing appropriate disinfectant, and to an in-line HEPA or equivalent filter.

This personnel will provide protection to the central building vacuum system or vacuum pump, as well as to the personnel who service this equipment. Inactivation of aspirated materials can be accomplished by placing sufficient chemical decontamination solution into the flask to kill the microorganisms as they are collected. Once inactivation occurs, liquid materials can be disposed of appropriately as noninfectious waste.



WWW.NUAIRE.COM